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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO
10/048,186	06/19/2002	James C Liao	06497-013002	2905
7590 06/30/2005			EXAMINER	
Fish & Richardson			PROUTY, REBECCA E	
225 Franklin Street Boston, MA 02110-2804			ART UNIT	PAPER NUMBER
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Please find below and/or attached an Office communication concerning this application or proceeding.

	Application No.	Applicant(s)				
	10/048,186	LIAO, JAMES C				
Office Action Summary	Examiner	Art Unit				
	Rebecca E. Prouty	1652				
The MAILING DATE of this communic Period for Reply	ation appears on the cover sheet w	vith the correspondence address				
A SHORTENED STATUTORY PERIOD FO THE MAILING DATE OF THIS COMMUNIC - Extensions of time may be available under the provisions of after SIX (6) MONTHS from the mailing date of this communication of the period for reply specified above is less than thirty (30). - If NO period for reply is specified above, the maximum statused in the period for reply within the set or extended period for reply within the set or exten	CATION. f 37 CFR 1.136(a). In no event, however, may a nication. days, a reply within the statutory minimum of thi atory period will apply and will expire SIX (6) MO ill, by statute, cause the application to become A	reply be timely filed rty (30) days will be considered timely. NTHS from the mailing date of this communication. BANDONED (35 U.S.C. § 133).				
Status						
1)⊠ Responsive to communication(s) filed	on <u>20 April 2005</u> .					
2a) ☐ This action is FINAL . 2t	o)⊠ This action is non-final.					
,	Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under <i>Ex parte Quayle</i> , 1935 C.D. 11, 453 O.G. 213.					
Disposition of Claims						
4)⊠ Claim(s) <u>1,5,11-13,17,21-24,37,38,40</u> 4a) Of the above claim(s) <u>11,22,23,37</u> 5)□ Claim(s) is/are allowed. 6)⊠ Claim(s) <u>1,5,13,17,21,24,40,41 and 4</u> 7)⊠ Claim(s) <u>12 and 55</u> is/are objected to. 8)□ Claim(s) are subject to restricti	.38,46-50,52-54 and 56-74 is/are 5 is/are rejected.					
Application Papers						
9)☐ The specification is objected to by the	Examiner.					
10)☐ The drawing(s) filed on is/are: a)☐ accepted or b)☐ objected to by the Examiner.						
Applicant may not request that any object	***	• •				
Replacement drawing sheet(s) including to the second of the second second in the second secon	·					
Priority under 35 U.S.C. § 119						
12) Acknowledgment is made of a claim for a) All b) Some * c) None of: 1. Certified copies of the priority d 2. Certified copies of the priority d 3. Copies of the certified copies of application from the Internation. * See the attached detailed Office action	ocuments have been received. ocuments have been received in a f the priority documents have been al Bureau (PCT Rule 17.2(a)).	Application No n received in this National Stage				
Attachment(s)						
1) Notice of References Cited (PTO-892)		Summary (PTO-413)				
 Notice of Draftsperson's Patent Drawing Review (PTo3) Information Disclosure Statement(s) (PTO-1449 or Paper No(s)/Mail Date 		(s)/Mail Date Informal Patent Application (PTO-152) 				

Application/Control Number: 10/048,186
Art Unit: 1652

A request for continued examination under 37 CFR 1.114, including the fee set forth in 37 CFR 1.17(e), was filed in this application after final rejection. Since this application is eligible for continued examination under 37 CFR 1.114, and the fee set forth in 37 CFR 1.17(e) has been timely paid, the finality of the previous Office action has been withdrawn pursuant to 37 CFR 1.114. Applicant's submission filed on 4/20/05 has been entered.

Claims 2-4, 6-10, 14-16, 18-20, 25-36, 39, 42-44 and 51 have been canceled. Claims 1, 5, 11-13, 17, 21-24, 37, 38, 40, 41, 45-50, 52 and newly presented claims 53-74 are still at issue and are present for examination.

Applicants' arguments filed on 4/20/05, have been fully considered and are deemed to be persuasive to overcome some of the rejections previously applied. Rejections and/or objections not reiterated from previous office actions are hereby withdrawn.

Applicant's election with traverse of Group I, Claims 1, 5, 11-13, 17, 21, 24, 37, 38, 40, 41, 45, 52 and new claims 53-57 and of *glnAp2* as promoter species and isopentenyl diphosphate isomerase (idi) as heterologous polypeptide in the response filed 12/5/03 is acknowledged. Applicants request rejoinder of method claims 22, 23, 46-50, and 58-74 and claims to the non-

Art Unit: 1652

elected species i.e., 11, 37, 38, 52-54, 56 and 57. However, The methods of Groups II (i.e., methods of producing an isoprenoid) and III (i.e., methods of producing lycopene) as defined in the lack of unity of 10/3/03 do not share any special technical feature with the products of Group I as many products within Group I cannot be used for the processes of Groups II and III. For example a glnL E. coli cell transformed with a polyketide synthase gene operably linked to the glnAp2 promoter which is clearly a product within the scope of Group I cannot be used for the methods of either of groups II or III. Furthermore the claims to the non-elected species within group I remain withdrawn as the linking generic claims are not allowable over the art (see the rejections herein).

Claims 11, 22, 23, 37-38, 46-50, 52-54, 56 and 57-74 are withdrawn from further consideration pursuant to CFR 1.142(b) as being drawn to a nonelected invention (Claims 22, 23, 46-50, and 58-74) or species (Claims 11, 37, 38, 52-54, 56 and 57), there being no allowable generic or linking claim. Claims 1, 5, 12, 13, 17, 21, 24, 40, 41, 45 and 55 are examined herein.

The disclosure is objected to because it contains an embedded hyperlink and/or other form of browser-executable code (see for example page 2 of the specification). Applicant is

Art Unit: 1652

required to delete the embedded hyperlink and/or other form of browser-executable code. See MPEP § 608.01.

Claims 13 and 41 are rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

Claim 13 is confusing in the recitation of "the host cell of claim 17 wherein the enzyme is ... 1-deoxyxylulose 5-phosphate synthase" because claim 17 from which claim 13 depends limits the enzyme to an enzyme that catalyzes biosynthesis of an isoprenoid but the product of 1-deoxyxylulose 5-phosphate synthase (i.e., 1-deoxyxylulose 5-phosphate) is not an isoprenoid.

Claim 41 is confusing in the recitation of "the host cell of claim 40 wherein the biosynthetic enzyme is ... 1-deoxyxylulose 5-phosphate synthase or phosphoenolpyruvate synthase" because claim 40 from which claim 41 depends limits the biosynthetic enzyme to an enzyme that catalyzes biosynthesis of an isoprenoid, polyketide or polyhydroxyalkanoate but the products of 1-deoxyxylulose 5-phosphate synthase (i.e., 1-deoxyxylulose 5-phosphate) and phosphoenolpyruvate synthase (i.e., PEP) are not an isoprenoid, polyketide or polyhydroxyalkanoate.

Application/Control Number: 10/048,186
Art Unit: 1652

Claims 1, 5, 17, 21, 24, 40, and 45 are rejected under 35 U.S.C. 112, first paragraph, because the specification, while being enabling for E. coli having an inactivating glnL mutation which are transformed with a nucleic acid encoding one or more of the enzymes isopentenyl diphosphate isomerase, geranylgeranyl diphosphate synthase, 1-deoxyxylulose 5-phosphate synthase and phosphoenol pyruvate synthase operably linked to the glnAp2 promoter, kits comprising an E. coli having a glnL mutation and a nucleic acid encoding the glnAp2 promoter or nucleic acid constructs therefore, does not reasonably provide enablement for E. coli having an inactivating glnL mutation which are transformed with a nucleic acid encoding any biosynthetic enzyme for the production of any isoprenoid, any polyketide or any polyhydroxyalkanoate. The specification does not enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and use the invention commensurate in scope with these claims.

These claims are so broad as to encompass any *E. coli* having an inactivating *glnL* mutation which are transformed with a nucleic acid encoding any biosynthetic enzyme for the production of any isoprenoid, any polyketide or any polyhydroxyalkanoate and the use of such cells for the production of any isoprenoid, any polyketide or any

Art Unit: 1652

polyhydroxyalkanoate. The scope of each of these claims is not commensurate with the enablement provided by the disclosure with regard to the extremely large number of genes necessary for the construction of the host cells broadly encompassed by the claims nor for the use of all such cells for the production of any isoprenoid, any polyketide or any polyhydroxyalkanoate. Isoprenoids, polyketides and polyhydroxyalkanoates each encompass an enormous family of highly complex compounds which are synthesized by highly complex biosynthetic pathways by an enormous number of different enzymes many of which are present in only a small number of microorganisms. Furthermore, wild type E. coli do not produce any polyketides or polyhydroxyalkanoates and produce only a very limited number of isoprenoids. Furthermore, while genes for the synthesis of some isoprenoids, polyketides and polyhydroxyalkanoates are provided by the specification and/or prior art, use of any combination thereof for the production of any isoprenoid, polyketide or polyhydroxyalkanoate is not a straightforward process involving only the transformation of a single gene into E. coli and expression of this gene therein. For many isoprenoids, polyketides and polyhydroxyalkanoates to be produced multiple genes are necessary only some of which may be available in the prior art, the biosynthetic pathways as well as competing

Art Unit: 1652

metabolic processes are not well defined, and all necessary precursors may not be present or may not be present in sufficient amounts. See for example the teachings of Kholsa et al. regarding the difficulties of expressing polyketides in E. Thus, the art provides little predictability of which coli. isoprenoids, polyketides and polyhydroxyalkanoates can be produced using the teaching of the specification. applicants have not provided sufficient guidance to enable one of ordinary skill in the art to make and use the claimed invention in a manner reasonably correlated with the scope of the claims broadly including any E. coli having an inactivating qlnL mutation which are transformed with a nucleic acid encoding any biosynthetic enzyme for the production of any isoprenoid, any polyketide or any polyhydroxyalkanoate and the use of such cells for the production of any isoprenoid, any polyketide or any polyhydroxyalkanoate. The scope of the claims must bear a reasonable correlation with the scope of enablement (In re Fisher, 166 USPQ 19 24 (CCPA 1970)). Without sufficient quidance, determination of the identity of bacterial cells having the desired biological characteristics is unpredictable and the experimentation left to those skilled in the art is unnecessarily, and improperly, extensive and undue. See In re Wands 858 F.2d 731, 8 USPQ2nd 1400 (Fed. Cir, 1988).

Art Unit: 1652

Claims 1, 5, 40, and 45 are rejected under 35 U.S.C. 103(a) as being unpatentable over either of Khosla et al. (US PG-PUBS 2002/0045220) or in view of Bock et al. (US Patent 5,830,692), McCleary et al. (Reference AL of applicant's PTO-1449), McCleary et al. (Reference AM of applicant's PTO-1449) and Haldiman et al. (Reference AK of applicant's PTO-1449) or Feng et al. (Reference AJ of applicant's PTO-1449).

Khosla et al. teach constructs for the recombinant expression of polyketide synthase modules in *E. coli* cells for the production of polyketides. The constructs of Khosla et al. comprise the polyketide synthase gene under the control of the inducible lac promoter. (see Fig 6). Khosla et al. further teach that the initial precursors of all polyketides include acetyl-CoA.

Bock teach that inducible promoters such as lac, tac, and trp promoters possess several disadvantages in relation to their use for industrial production. These are that the repressors and inducers necessary for use of these promoters are expensive and difficult to handle, particularly when they are metabolizable substances (such as lactose and tryptophan), and cannot be induced completely when the repressor is present in molar excess. (see columns 1-2).

Art Unit: 1652

McCleary (AK) and McCleary (AP) teach that acetyl phosphate may act a global regulatory signal in *E. coli* responsible for the activation of a wide range or response regulators of two-component systems, including the *glnAp2* promoter, in the absence of their cognate histidine kinase (i.e., the *ntrB* gene product in the case of *glnAp2*). They further teach that acetyl-phosphate levels in bacteria correlate with the amount of acetyl-CoA produced and is present at high levels whenever glycolytic intermediates accumulate.

Haldiman et al. and Feng et al. each teach *E. coli* twocomponent system promoters (the *VanH* promoter in Haldiman et al.
and the *glnAp2* promoter in Feng et al.) which are activated by a
response regulator protein (*VanR* in Haldiman et al. and *NtrC* in
Feng et al.) and acetyl phosphate in the absence of the
corresponding histidine kinases (*VanS* in Haldiman et al. and *NtrB* in Feng et al.) and in the presence or absence of nitrogen.

As inducers (IPTG) for promoters such as *lac* used by Khosla et al. are expensive and have disadvantages as taught by Bock, it would have been obvious to one of ordinary skill in the art to link the production of the polyketide synthase to the presence of a metabolite in the cell which signals that significant amounts of the precursors for polyketide biosynthesis are present. McCleary et al. (1993) teach that

Art Unit: 1652

acetyl phosphate accumulation occurs under these conditions. Therefore, it would have been obvious to one of ordinary skill in the art to replace the lac promoters in the constructs of Khosla et al. with a promoter which is induced by high acetyl phosphate levels. As McCleary et al. (AK and AP) teach that acetyl-phosphate levels correlate with the amount of acetyl-CoA produced, it would have been obvious to one of ordinary skill in the art to link the polyketide synthase genes of Khosla et al. to the acetyl-phosphate regulated promoters taught by Haldiman et al. or Feng et al. and express these constructs in E. coli cells which lack the cognate histidine kinases such that the response regulators which activate transcription from these promoters are activated by acetyl phosphate. Furthermore, it would have been obvious to one of ordinary skill in the art to put the cells and vectors necessary for production of high levels of polyketides together in a kit for easy handling.

Claims 12 and 55 are objected to as being dependent upon a rejected base claim, but would be allowable if rewritten in independent form including all of the limitations of the base claim and any intervening claims.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Rebecca E. Prouty whose telephone number is 571-272-0937. The examiner can normally be reached on Tuesday-Friday from 8 AM to 5 PM. The examiner can also be reached on alternate Mondays

Art Unit: 1652

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Ponnathapura Achutamurthy, can be reached at (571) 272-0928. The fax phone number for this Group is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see http://pair-direct.uspto.gov. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

Rebecca Prouty Primary Examiner

Art Unit 1652